## INTRODUCTION

BTK inhibitors have revolutionized the management of chronic lymphocytic leukemia (CLL) and other B cell malignancies. However, acquired resistant mutation of BTK develops.

Mutations at Cysteine 481 (C481) result in resistance to covalent BTK inhibitors (BTKi). Non-covalent BTKis, such as pirtobrutinib, can overcome C481 mutations but are susceptible to other BTK mutations like V416L, T474I, and L528W.

Recently, a BTK mutation A428D was reported to confer resistance from the BGB-16673 trial and was also modeled to disrupt the binding of BTK to another BTK-degrader, NX-2127 [1]. This raises the question of whether A428D insensitivity is compound specific or common to BTK degraders and inhibitors currently in development and market. This study investigates the impact of BTK A428D on the response to approved BTK inhibitors and multiple degraders via preclinical models.

## METHOD

TMD8 cells overexpressing BTK-wildtype, C481S, T474I, A428D, V416L, M437R mutations were generated by lentivirus transduction and utilized to evaluate their sensitivity to individual BTK degrader or inhibitor via *in vitro* CTG assay.

TMD8 BTK-A428D cell lines, which were generated by CRISPR-Cas9 mediated in situ gene editing, were used for BTK degradation examination by ELISA, BCR signalling evaluation by Western blot, and *in vivo* tumour growth inhibition (TGI) assessment via xenograft models.

## REFERENCES

[1] Wong, R.L., Choi, M.Y., Wang, HY. et al. Mutation in Bruton Tyrosine Kinase (BTK) A428D confers resistance To BTK-degrader therapy in chronic lymphocytic leukemia. Leukemia 38, 1818–1821 (2024). https://doi.org/10.1038/s41375-024-02317-4

### Figure 2. TMD8 cells with BTK-A428D knock in (KI) mutation confirmed the resistance to BTK inhibitors and degraders TMD8 BTK-Wildtyp TMD8 BTK-A428D KI #117

IC50 (nM) **TMD8** Cells BGB-16673 NX- NX-5948 Abbv-Zanu Ibru **BTK-WT** 4.058 15.87 BTK-A428D KI 35# BTK-A428D KI 117# >10.000

Two TMD8 BTK-A428D cell lines generated by CRISPR-Cas9 mediated *in situ* mutation were treated by BTK degraders and inhibitors to confirm their response to different drugs. A-B). In parental TMD8 cells expressing wildtype BTK, cellular growth was efficiently inhibited by both BTK degraders and inhibitors; **C-F).** Neither four BTK degraders, nor the four tested BTK inhibitors could efficiently inhibit the growth of two TMD8 BTK-A428D knock in cell clones (Clone #35 and Clone #117), confirming their resistance to BTK-A428D mutations observed in OE cells in Figure 1; G). A table showing the number of half maximum inhibition concentration (IC50) of different compounds from A to F.

# BTK-A428D is a Cross-resistant Mutation to Both BTK Inhibitors and Degraders

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# RESULTS

## Figure 1. BTK-A428D is a cross resistant mutation to BTK inhibitors and degraders

Cell proliferation IC50 in TMD8 OE



474I, A428D, V416L, M437R mutations were treated different BTK degrader or inhibitors, and the cell viabilit

Abbv-Compound 1 is example 1 in WO 2023183811

Among the four tested BTK degraders, which are BGB-16673, NX-2127, NX-5948 and Abbv-Compound 1: 1) BGB-16673 and NX-5948 potently inhibited the growth of TMD8 cells overexpressing BTK-C481S, T474I, V416L, M437R mutations measured by in vitro CTG assay; 2) BGB-16673 is more potent than Abbv-Compound 1 against BTK-T474I, -V416L, and -M437R overexpressing TMD8 cells; 3) NX-2127 is generally less potent in all the mutations. However, TMD8 cells overexpressing BTK-A428D are resistant to all tested BTK degraders and inhibitors (ibrutinib, acalabrutinib, zanubrutinib, pirtobrutinib).



# CONCLUSIONS

BTK-A428D is a mutation that exhibits cross-resistance to multiple BTK degraders (BGB-16673, NX-2127, NX-5948, Abbv-Compound) 1) and inhibitors (ibrutinib, acalabrutinib, zanubrutinib, and pirtobrutinib) both *in vitro* and *in vivo*, as supported by evidence from multiple aspects.



BTK degradation were examined by ELISA and western blot assay in TMD8 wildtype and BTK-A428D knock-in cells. A). BGB-16673, NX-5948, and Abbv-Compound 1 potently degraded wildtype BTK in TMD8 cells, NX-2127 also efficiently degraded wildtype BTK, but showed lower potency compared with other degraders. B). None of the BTK degraders showed significant degradation on BTK-A428D protein, even at 1,000 nM. C). A table shows the half degradation concentration (DC50) in figure A, B. **D-E).** TMD8 wildtype or BTK-A428D KI cells were treated by different compounds at 100nM for 24h, then protein lysates were used for western blot. Significant BTK degradation was only observed by different BTK degraders in wildtype cells, and the degradation was blocked in BTK-A428D KI cells. As expected, no BTK degradation was observed for BTK inhibitors.

## Figure 3. BTK degradation by degraders is blocked by A428D mutation



## Figure 4. BTK-A428D is a kinase-impaired mutation, but maintains BCR downstream signaling

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		Wildtype			A428D-#35				A428D-#117				
1mM PV (min)	0	5	10	20	0	5	10	20	0	5	10	20	
p-BTK(Y223)		-	-	-									
ВТК	•	••		-	-	-	-	-	-	-			
p-PLCγ2 (Y1217)		•		-		-	-	-		-	-	_	
PLCγ2	-		-	-	-	-	-	-	-	-	-	-	
p-AKT (S473)		•	-	-	•	-	-	-		-	-	-	
AKT	-			-	-	-	-	-	-	-		-	
GAPDH	-			-	_	_	_	_	_	_		-	

TMD8 wildtype and two BTK-A428D *in situ* mutated clones (Clone #35 and Clone #117) were used to examine BTK kinase activity with or without 1mM pervanadate (PV) stimulation. A time-dependent increase of phosphorylation signal at BTK-Y223, PLCy2 -Y1217, and AKT-S473 were observed in TMD8 BTK-wildtype cells, suggesting an intact kinase activity of BTK and its downstream signals. However, no phosphorylation of BTK-Y223 can be observed in BTK-A428D cells, indicating A428D is a kinase impaired mutation. Consistently, phosphorylation at PLC<sub>2</sub> is also significantly abrogated. Phosphorylation at AKT-S473 is less disrupted, suggesting the BCR downstream signaling retains in mutant.



## Figure 5. BTK inhibitors and degraders are ineffective in inhibiting BTK downstream signaling in BTK-A428D mutated cells

Α	TMD8 BTK-Wildtype						TMD8 BTK-Wildtype							
1mM PV, 20min	- +	- ++				- +		+						
Compounds	vehicle	BGB-16673	NX-2127	NX-5948	Abbv-Cmpd1	Vehicle	Zar	าน	lbr	'n	Acala	Pirto		
Concentration (nM)		100 10 1	100 10 1	100 10 1	100 10 1		100 1	01	100 <sup>·</sup>	10 1	100 10 1	100 10 1		
p-BTK(Y223)				-		-		-		-				
ВТК														
p-PLCγ2(Y1217)	-					-	-			-				
PLCy2														
p-STAT3(Y705)														
STAT3														
GAPDH														
В		-			<b>Z</b> 1									
	WT	I		-A428D P		WT			/ID8		-A428D K			
1mM PV, 20min	+ -		•	+		+ -					+			
Compounds	vehicle	BGB-1667	3 NX-212	7 NX-5948	Abbv-Cmpd 1	vehicle	, <u>z</u>	anu		bru	Acala	Pirto		
Concentration (nM)		100 10 1	100 10 1	100 10 1	100 10 1		100	10 1	100	) 10	1 100 10	1 100 10 1		
p-BTK(Y223)	-					-								
ВТК														
p-PLCγ2(Y1217)	-								-					
PLCy2									-					
p-STAT3(Y705)									-					
STAT3														
GAPDH														

TMD8 wildtype and BTK-A428D knock-in cells were treated with compounds at indicated concentrations for 24 hours, then cells were stimulated with 1mM PV for 20min. Cell lysates were used for western blot. A). BTK was dose dependently degraded by BTK degraders (BGB-16673, NX-5948, NX-2127, Abbv-Compound 1) but not BTK inhibitors (zanu, ibru, acala, pirto). The phosphorylation of BTK, PLCγ2 and STAT3 were dose dependently inhibited by both BTK degraders and inhibitors, suggesting efficient inhibition of BTK and its downstream signal by both compounds. B). No obvious degradation of BTK-A428D was observed by different degraders, indicating the mutant is resistant to all BTK degraders. No BTK-Y223 phosphorylation can be observed, confirming it a kinase impaired mutation. None of the compounds significantly inhibited the phosphorylation of PLCγ2 and STAT3, suggesting they are ineffective in inhibiting downstream BCR signals in A428D mutant cells

### Figure 6. BTK inhibitors and degraders showed no obvious anti-tumor activity in BTK-A428D xenografts



A-B). No anti-tumor activity was observed for both BTK degraders and inhibitors in TMD8 BTK-A428D xenografts in vivo. C-D). Compounds showed significant anti-tumor activity in wildtype xenografts

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